

# CLINICAL IMPACT OF THE EUROPEAN MYELOMA NETWORK GUIDELINES FOR MULTIPARAMETRIC FLOW CYTOMETRY (MFC) ANALYSIS OF MYELOMA PLASMA CELLS - REPORT FROM THE NMSG #9/99 PROJECT

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**Background** Recently, the European Myeloma Network (EMN) has published guidelines for MFC in MM, which we have applied and here we present results from analysis of 280 bone marrow (BM) samples from teen Nordic centres that support a diagnostic, prognostic and predictive impact in clinical practice.

**Design and Methods** This registration study were planned to include a minimum of 100 diagnostic samples analysed in accordance with the EMN guidelines for MFC with the aims to evaluate the number and the phenotypic profile of myeloma plasma cells (PC) and the impact of minimal residual disease. Samples were analysed as described previously for identification of myeloma end stage CD45/CD38++ PC. Registration included relative numbers of events and calculated mean fluorescence intensity (MFI) for B cell membrane antigens and auto-fluorescence within the PC gate. All MFI ratios were calculated for cluster analysis performed with software by the hierarchical ordering of Pearson's coefficients method.

**Results** First, the median MFC-PC number was 15% and LM-PC 35% for corresponding samples significantly correlated (Figure 1a & b). The difference between MFC- and LM-PC numbers *at diagnosis* was significantly influenced only by the CD56 MFI on PC, indicating that homophile cell-cell binding may explain the difference. At least 10% plasma cells in bone marrow aspirate counted by light microscopy is a diagnostic demand and therefore we studied the variation of the MFC-PC number in the planned 80 MM and 20 MGUS patients as illustrated in Figure 1a. The distributions indicate that a cut off value detected by MFC of 1% PC (Figure 2a) discriminate MM disease and premalignant MGUS samples with a sensitivity of 0.83 (95% C.I. 0.75-0.92) and specificity of 0.53 (95% C.I. 0.30-0.75) respectively. Such a discriminatory value is based on the dataset shown in Figure 2b from analysis of 67 non diagnostic BM samples received during the study period. These samples were aspirated due to a clinical need for PC enumeration for evaluation of treatment outcome. These data therefore reflects daily decision taking for BM analysis monitoring patients influenced by symptoms, organ involvement or the level of M component and seem to divide patients in two clear groups by MFC-PC number of 1%. These results are in support for inclusion of plasma cell number by MFC into the diagnostic triad for MM – but needs to be prospectively validated.

Second, hierarchical unsupervised clustering of MFI revealed a patient cluster I with high membrane intensity for CD56, CD38, CD45, SSC, FSC, and low for CD28, CD19 and a patient cluster II, (N = 50) with low intensity of CD56, CD38, CD45, SSC, FSC and high for CD28, CD19 (Figure 3a). The *prognostic impact* of such clusters revealed a median survival of 39 and 19 months respectively (P=0.02) (Figure 3b).

Third, enumeration of minimal residual disease in bone marrow aspirates from 16 high dose melphalan treated patients revealed a reduction of MFC-PC from median 1.7% at diagnosis to 0.22% and 0.16% following 1st and 2nd transplantation (Figure 4a). A further decreased level 2-3 months after the 2<sup>nd</sup> HDT *predicted superior* EFS (Figure 4b).

**Conclusions** The guidelines for MFC in MM do produce diagnostic, prognostic and predictive information useful in clinical practice, which however needs to be prospectively validated as planned within EMN.

**Reference** Report of the European Myeloma Network on multiparametric flow cytometry in multiple myeloma and related disorders. Rawstron AC, Orfao A, Beksac M, Bezdicikova L, Brooimans RA, Bumbea H, Dalva K, Fuhler G, Gratama J, Hose D, Kovarova L, Lioznov M, Mateo G, Morilla R, Mylin AK, Omedé P, Pellat Deceunynck C, Perez Andres M, Petrucci M, Ruggeri M, Rymkiewicz G, Schmitz A, Schreder M, Seynaeve C, Spacek M, de Tute RM, Van Valckenborgh E, Weston-Bell N, Owen RG, San Miguel JF, Sonneveld P, Johnsen HE; European Myeloma Network. *Haematologica*. 2008 Mar;93(3):431-8.

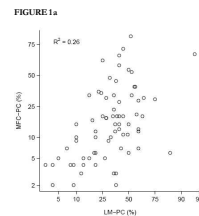


Figure 1a Legend: Correlation of light microscopically (LM-PC) and all plasma cell percentage by MFC (MFC-PC) and by light microscopy (LM-PC) significantly correlated data points (grey with MFC-PC number) from 280 patients. LM-PC number corresponding to the LM-PC gate. LM-PC number corresponding to the LM-PC gate. LM-PC number corresponding to the LM-PC gate.

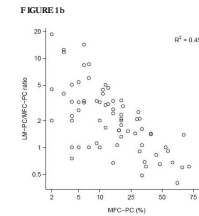


Figure 1b Legend: Correlation of LM-PC/MFC-PC ratio and the plasma cell percentage enumerated by MFC. Correlation coefficient. No correlation was identified when LM-PC is compared to plasma cell percentage enumerated by LM (not shown).

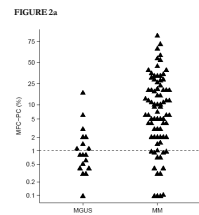


Figure 2a Legend: Analytic plot showing the variation of the MFC-PC number in 20 MGUS and 80 MM patients. Median MFC-PC is 1.5% and 2.5% respectively. The horizontal dashed line indicates the cut-off value detected by MFC of 1% PC. The plot shows that MM samples have significantly higher MFC-PC numbers compared to MGUS samples. The plot shows that MM samples have significantly higher MFC-PC numbers compared to MGUS samples.

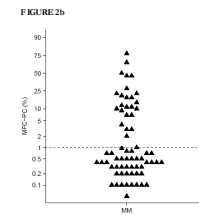


Figure 2b Legend: Analytic plot showing MFC-PC numbers in 67 non-diagnostic MM samples. The plot shows that MM samples have significantly higher MFC-PC numbers compared to MGUS samples. The plot shows that MM samples have significantly higher MFC-PC numbers compared to MGUS samples.

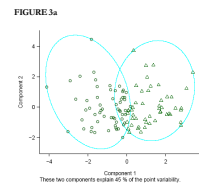


Figure 3a Legend: Hierarchical clustering of MFI values. The plot shows two distinct clusters of patients based on MFI values. The plot shows two distinct clusters of patients based on MFI values.

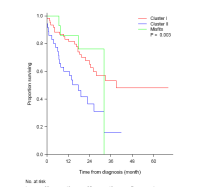


Figure 3b Legend: Kaplan-Meier survival plot showing survival probability over time (months) for two patient clusters. The plot shows that patients in cluster I have a significantly better survival outcome compared to patients in cluster II.

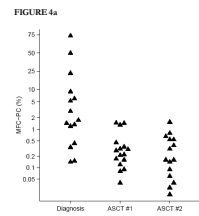


Figure 4a Legend: Enumeration of minimal residual disease by the MFC-PC number. The plot shows that MFC-PC percentages are significantly lower after the first and second ASCT compared to the diagnosis. The plot shows that MFC-PC percentages are significantly lower after the first and second ASCT compared to the diagnosis.

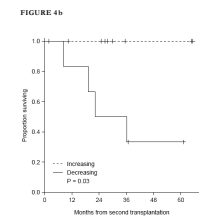


Figure 4b Legend: A decreased EFS level following 2nd HDT. The plot shows that patients with high MFC-PC percentages at diagnosis have a significantly better EFS outcome compared to patients with low MFC-PC percentages at diagnosis. The plot shows that patients with high MFC-PC percentages at diagnosis have a significantly better EFS outcome compared to patients with low MFC-PC percentages at diagnosis.